

Antimicrobial, Antioxidant and Cytotoxic Activity of Green Synthesized Copper Nanoparticle of *Parthenium hysterophorus* L.



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ABSTRACT: Natural products are valuable and well known for their biological activities. In the current scenario, the research and analysis of plant leaf extract and nanoparticles synthesis with their biological activities has been expended significantly. For the synthesis of nanoparticles, Copper is the preferred metal among other metals due to its reported use in medical field as antimicrobial agents and its lethality. The aim of the present work is to assess antimicrobial and antioxidant activities of the methanolic leaf extracts and aqueous leaf extract mediated copper nanoparticles of *Parthenium hysterophorus*. Study also revealed the comparative analysis between methanolic leaf extracts and copper nanoparticles. Synthesis of copper nanoparticles (CuNPs) was confirmed by the change of color of aqueous extract, which were further confirmed by using UV-Vis spectrophotometer. The results indicated that copper nanoparticles have great antimicrobial activity against *Bacillus subtilis*, *Staphylococcus aureus*, *Candida albicans* and *Aspergillus flavus* rather than methanolic leaf extract. These studies confirmed that copper nanoparticles are capable to rendering high antimicrobial efficacy and hence has a great potential in the development of antimicrobial agents. Based on the DPPH, copper nanoparticles found to be good antioxidant when compared to methanolic leaf extract. Furthermore cells viability assay was also done against copper nanoparticles. These results concluded that copper nanoparticles are good source of therapeutic agent and applications of copper nanoparticles based on these findings may lead to valuable discoveries in various fields such as medical devices and antimicrobial systems.

KEYWORDS: *Parthenium hysterophorus*, Copper nanoparticles, antimicrobial activity, antioxidant activity, cell viability assay

1. INTRODUCTION

Parthenium hysterophorus L. (Asteraceae) is an aggressive annual herbaceous plant also known as Congress grass or Carrot weed and native to the Tropical America. It is now widely distributed in a number of tropical and sub-tropical countries (Gnanavel *et al.*, 2013). In India, the weed has achieved major weed status already. It is very common along with the road sides, around the agricultural fields and on waste lands and considered as a noxious weed because of its prolific seed production and fast spreading ability, allelopathic effect on other plants, strong competitiveness with crops and health hazard to human as well as animals (Reviewed by Kaur *et al.*, 2014). Phytochemistry of plant showed various toxic compounds such as Parthenin, Coronopilin, 2b-hydroxycoronophilin, Tetraneurin-A (Patel. Seema, 2011). The infestation of the weed causes yield losses up to 40% in several crops and reduces forage production by up to 90%. The rapid spread of *Parthenium* in India would be a bigger risk to the expansion and sustainable production of many crops (Sushilkumar, 2014).

Control of *Parthenium* is therefore crucial to boost the productivity of agricultural crops in the country. It is reported that for better management of the weeds, knowing the habitat, morphology and biology of the weeds are also important. It can be kept control by enhancing its use in different aspects. Exploitation of the weed for the beneficial use should be promoted. The increased utilization of *P. hysterophorus* as insecticide, pesticide, and composite, raw material for biofuel and enzyme production can change the weed from a curse to a boon for civilization (Kaur *et al.*, 2014). Antimicrobial and anticancerous nature of *P. hysterophorus* was reported already (Tapwal *et al.*, 2011; Kumar *et al.*, 2013).

Various researches in the field of antimicrobial development using nanoparticals are gaining interest as they can be used as an alternative to antibiotic (Ge *et al.*, 2014). For the biosynthesis of nanoparticles, plant mediated approach is used now days which is a cost-effective and environmentally friendly alternative to chemical and physical methods (Niraimathi *et al.*, 2013).

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Considering the vast potentiality of plants as a source for drug and its usefulness in the biosynthesis of metal nanoparticles, a systematic investigation was undertaken to screen a widespread available flora *P. hysterophorus*.

Natural products such as plant extracts provide unlimited opportunities for new drug discoveries (Sasidharan *et al.*, 2011). Recent evidences from the pharmaceutical companies show that it still represent an extremely valuable chemical entities that can be used for the treatment of some complex diseases (Chin *et al.*, 2006). These medicinal plants can be rich in phenolic compounds, alkaloids, diterpenoids, steroids and other compounds which inhibit the development of various microorganisms (Ranjitham *et al.*, 2013). Besides this, phytochemicals in the plants extracts can act as reducing and capping agents in the reduction of metal ions to metal nanoparticles (Swarnalatha *et al.*, 2013) and thus have found widespread use in the biosynthesis of metal nanoparticles which can be used in drug delivery (Doane and Burda, 2012), biosensing (Bedford *et al.*, 2012), catalysis (An and Somerjai, 2012). Recently, the biosynthesis of metal nanoparticles using plants extracts has received considerable attention as a suitable alternative to existing synthesis of nanoparticles by chemical procedure and physical methods (Song and Kim, 2009; Chandran *et al.*, 2006). It is also very cost effective (Sujitha and Kannan, 2013; Geethalakshmi and Sarada, 2012) and thereby it can be used as an economic and valuable substitute for the large scale production of metal nanoparticles.

Nanoparticles are small-sized materials of size range between 1-100nm. These nanoparticles are more active and exhibited unexpected properties because of high surface to volume ratio and quantum size effect (Luo *et al.*, 2005; Monga and pal, 2015). Furthermore for the agriculture purpose, nanoparticles of colloidal particulate include the size range between 10-1000 nm. (Nakache *et al.*, 1999). Copper is most widely used material in the world; it has various applications among a variety of metal particles such as gold, silver, iron, palladium, zinc etc. such as their electrical, optical, catalytic, biomedical and antimicrobial. The copper is highly toxic to microorganism such as bacteria (*Escherichia Coli*, *Pseudomonas aeruginosa*) whereas non-toxic to animal cells, due to these phenomena it is considered to be an effective antimicrobial metal (Pawar *et al.*, 2014). Many researchers have reported the biosynthesis of metal nanoparticles by using various plants extracts (He *et al.*, 2013, Niraimathi *et al.*, 2013).

Considering the vast potentiality of plants as a source for drug and its usefulness in the biosynthesis of metal nanoparticles, a systematic investigation was undertaken to screen a local flora, *Parthenium hysterophorus*.

Justification of the Work *P. hysterophorus* is most obnoxious exotic weed and widely spreading in all agro-climatic conditions of India; affecting agriculture, livestock health, human health, soil conditions, and native biodiversity. Concurrently it is neither traditionally nor commercially used in India. Since it is exerting broad spectrum allopathic effect over native plant biodiversity, therefore, this weed can kept control by enhancing its beneficial use should be promoted so that increased utilization can change the weed from a curse to a boon for civilization.

The present work is therefore justified in terms of the:

1. Addition of information regarding antimicrobial nature of *P. hysterophorus*.

The aim of the study is to assess the antimicrobial, antioxidant activity and cytotoxic effect of methanolic extract as well as copper nanoparticles from leaves of experimental plant *P. hysterophorus*.

2. MATERIAL & METHODS

2.1 Chemicals and Reagents

All chemicals and reagents used in the experiment were analytical grade. Copper sulfate (CuSO_4) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) were purchased from SRL, and used without further purification. For the preparation of chemicals and reagents Milli-Q water was used during the experiments. The bacterial and fungal strains were obtained from ABPL, Lucknow, Uttar Pradesh. Bacterial and fungal culture namely *Bacillus subtilis*, *Staphylococcus aureus*, *Aspergillus flavus* and *Candida albicans* were maintained in the respective medium.

2.2 Collection of Plant Material Collection of Plant material and Preparation of sample

Diseased free leaves of experimental plant PH will be collected from local areas of Allahabad, (U.P.) during the month of December. The raw material (leaves) will be carefully washed in order to remove dirt, dust and residues of the pesticide spray and dried under shade for 24hrs. After that material will be dried at 55°C in hot air oven for 4-5 days or until constant weight will be obtained and then ground in course powdered by electric grinding machine. Powdered material will be further passed from sieve #20 and stored in air tight containers/desiccators until used. Powdered plant material will be weighed and used for the preparation of aqueous and solvent extracts.

2.3 Preparation of organic and inorganic extract

The dried and powdered plant material (10gms) was extracted successively with 30mL of methanol (organic) kept in refrigerator

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for maceration and filtered with Whatman filter paper. The obtained extract was left in oven (60°C) to evaporate. The dried extract was weighed (1gm.) and used for experiments. For the aqueous leaf extract (inorganic) (10gm) was weighed and added to 100mL of Milli-Q water to make infusion filtered through Whatman No. 1 filter to obtain aqueous leaf extract used for synthesis of copper nanoparticles.

Synthesis and Characterization of Copper Nanoparticles:

Approximately 1mM copper sulfate ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$) was used for the synthesis of *P. hysterophorus*-mediated copper nanoparticles synthesis. The syntheses of copper nanoparticles (CuNPs) 100 mL of aqueous leaf broth were added to 250 mL of aqueous copper sulfate solution in stirring condition. The concentration that gave the best results was further tested at room temperature and various time intervals. The reaction mixture was filtered through whatman filter paper for the extraction of Li-CuNPs. The residue was further purified by washing three times from distilled water to remove the water soluble biomolecules that were present. From this experiment, the best synthesis condition was established, and this condition was employed for the mass production of nanoparticles for further characterization and evaluation of their biological activities.

Characterization of synthesized nanoparticles:

The plant leaf mediated copper nanoparticles were characterized by commonly two method namely visual inspection and UV-Vis spectra analysis.

Visual Inspection: The bio-reduction of copper nanoparticles using aqueous extract of leaf observed by change of the aqueous leaf extracts color and exhibit turbid indication.

2.4 UV-Vis spectra analysis

The reduction of copper sulfate to copper using aqueous extract was monitored by measuring the UV-Vis spectrum at the range of 250-750 nm. The reaction mixture was initially diluted and aliquation was done with milli-Q water. Later the measurements of turbidity were recorded on double beam spectrophotometer (Labornics-Model CT-2800) at time interval of 3hrs and 24hrs.

2.5 Evaluating Of Biological Activities

Antibacterial Screening

The initial determination of the antibacterial activity, agar well diffusion was performed (**Naz and Bano, 2013**). The antimicrobials present in the plant extract and copper nanoparticles are allowed to diffuse out into the medium and interacted in a plate freshly seeded with the test bacteria on LB agar medium. The resulting zone of inhibition measured in millimeters.

The confirmatory assay called as antibacterial kill-kinetic assay was used to determine the bactericidal activity of methanolic leaf extract and copper nanoparticles. According to standard guide for assessment of antimicrobial activity using growth kinetics procedure with some modifications (**Antimicrobial Susceptibility Testing Method, 2008**), time- kill kinetics of the sample on test bacteria were carried out to assess the killing rate of the extract within a given contact time.

2.6 Antifungal Analysis

Antifungal screening was carried out using disc diffusion method **Kirby-Bauer (1966)**. Petri plates were prepared with 20mL of sterile Potato-Dextrose agar (PDA). The test culture were swabbed on the top of the solidified media and allowed to dry for 15min. Different amount (5µL, 10µL, 15µL, 20µL, 25µL and 50µL from 100µg/mL) of methanolic leaf extract and CuNPs. The loaded discs were placed on the surface of the medium and left for 30 min at room temperature. The plates were incubated for 48-72 h at 28°C for fungal growth. Zone of inhibition were recorded in millimeters, the fungal strains were *Aspergillus flavus* and *Candida albicans*. Streptomycin used as positive control drug and DMSO was used as negative control for antifungal screening. Experimental results were given as mean ± S.D. of the two parallel measurements.

2.7 Antioxidant Activity

The antioxidant activity of the extracts was evaluated by DPPH radical scavenging assay which was originally described by **Blois (1958)**. In brief, 0.1mM DPPH in 95% ethanol was prepared. This solution (1mL) was added to 3 mL of sample (methanolic leaf extract and copper nanoparticles) in ethanol at a concentration (100µg/mL). The mixture was shaken vigorously and allowed to stand at room temperature for 30 min. Then, absorbance was measured at 517 nm by using UV-Vis spectrophotometer. Reference standard compound being used was ascorbic acid and experiment was done in triplicate (**Patel and Patel, 2011**). Lower absorbance of the reaction mixture indicated higher free radical scavenging activity (**Koleva et al., 2002**). The percentage DPPH scavenging effect was calculated by using following equation:

$$\text{DPPH scavenging effect (\%)} \text{ or Percent inhibition} = \frac{A_0 - A_1}{A_0} \times 100.$$

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Where A_0 was the Absorbance of control reaction and A_1 was the Absorbance in presence of test or standard sample (Achola *et al.*, 1998).

2.8 Cell Viability Assay

In order to assess the activity of copper nanoparticles against mouse macrophage cell line J-774A.1, was used. The cell viability was determined with 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay (Mosmann, 1983) and some modification (Ahmad *et al.*, 2011; Loisean *et al.*, 2011). Exponentially growing cells J774.A-1 (Mouse macrophage) (1×10^4 cells /100 μ l/well) are incubated in RPMI-1640 containing 10% FBS with different sample concentrations (30-100 μ g/ml) for 72 hours and are incubated at 37°C in a humidified mixture of CO₂ and 95 % air in an incubator. Stock solutions of compounds are initially dissolved in DMSO and further diluted two fold with fresh complete medium. After incubation, 25 μ l of MTT reagent (5mg/ml) in PBS medium, followed by syringe filtration, was added to each well and incubated at 37 °c for 2 hours. At the end of the incubation period, the supernatant is removed by tilting plate completely without disturbing cell layer and 150 μ l of pure DMSO are added to each well. After 15 min. of shaking, the readings are recorded as absorbance at 544 nm on a micro plate reader. The cytotoxic effect are expressed as 50% cytotoxic concentration, i.e., as the concentration of a compound which provoked a 50% reduction in cell viability compared to cell in culture medium alone.

2.9 Biochemical Studies

The prepared leaf sample of test plant was used for the determination of total carbohydrate by colorimetric method using anthrone, (Morris, 1948). The water soluble protein was estimated by following the method of Lowry *et al.* (1951).

3. RESULTS AND DISCUSSION

3.1 Characterization of Copper Nanoparticles

3.1.1 Visual Inspection

During the biosynthesis, formation of nanoparticles was indicated by the change in color of the mixture (copper sulphate and leaf extract). After reaction, turbidity exhibited and the reaction mixture changed its color from intense green to brown color indicating the synthesis of Cu nanoparticles due the reduction of cupric (Cu⁺) ions and due to the excitation of surface plasmon vibration in metal nanoparticles.



Figure 1: Visual Inspection of CuNPs

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3.1.2 UV-Vis Spectra Analysis

UV-Visible absorption spectrum is the preliminary characterization to know the absorbance of synthesized nanoparticles. The results obtained from UV-Vis spectra analysis of the nanoparticles sample. It is the most important method of analysis to detect the Surface Plasmon Resonance property of CuNPs (Curtis *et al.*, 1998). Absorption spectrum of the prepared copper nanoparticles were taken in different time intervals up to 24hrs from the reaction mixture, in the range of 250-750nm by using double beam Spectrophotometer (Labronics-Model CT-2800). The CuNPs formation was confirmed from the peak around 450 to 550 nm. The peak value was found to be gradually decreased with increase in particle size. Copper SPR effects decrease with the time because of the oxidation of the synthesized copper nanoparticles (Tilaki *et al.*, 2007).

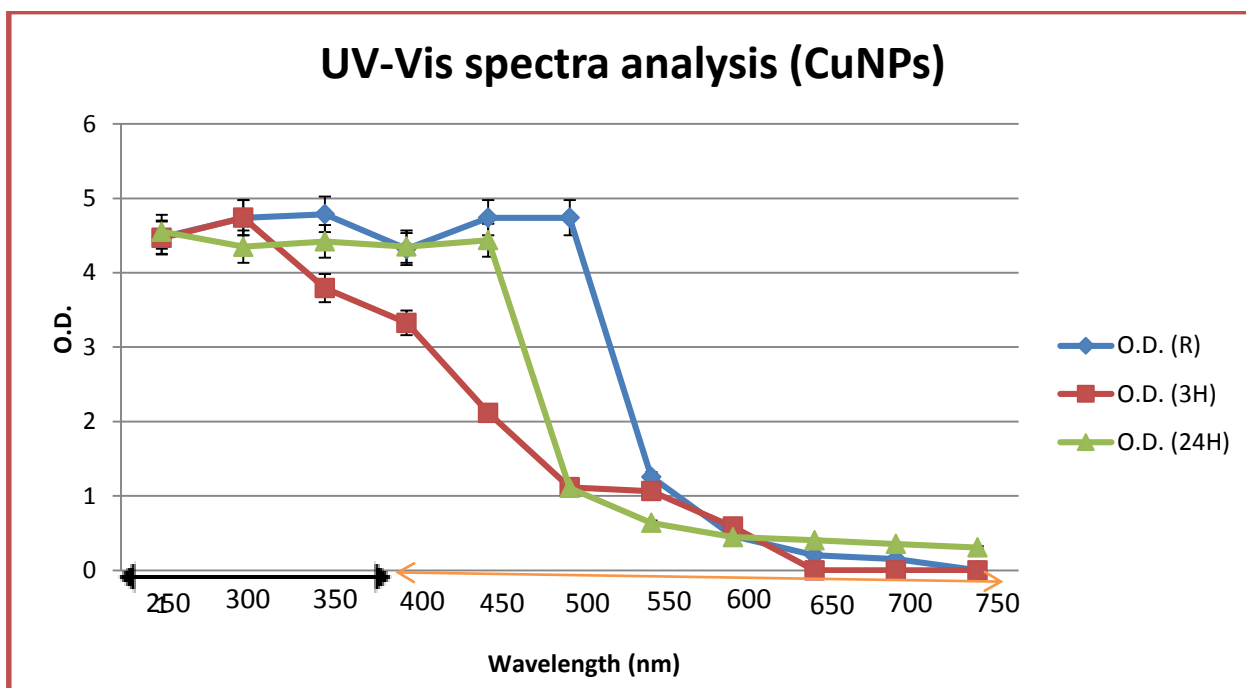


Figure 2: UV-Vis absorption spectra analysis

Where, R= Reference (Aqueous leaf extract)

↔ = Indicating ultraviolet range (250-380nm)

↔ = Indicating visible range (390-750nm)

3.2 Antibacterial Activity

The antibacterial results of the nanomaterial were compared to the metal free *P. hysterophorus* methanolic leaf extract. At a concentration of 100µg/mL in DMSO (as a diluent), the LI-CuNPs were active to both gram positive bacterial strains (*Bacillus subtilis* & *Staphylococcus*) used for the antibacterial screening. The nanoparticles showed greater antibacterial activity compared to plant extract of methanolic leaf extract of experimental plant. The enhanced antibacterial activity of the CuNPs could be attributed to their liposolubility. This is because, according to Overtone's concept of cell permeability, the lipid membrane that surrounds the microbial cell favours the passage of only the lipid-soluble substances. Hence lipophilicity is a major property for antibiotics. In addition, Tweedy's chelation theory proposed that on chelation of metal ions by ligands (constituent of plant extract) causes the polarity of the metal ion to be reduced to a greater extent due to the orbital overlap (between the metal and ligand orbitals) and partial sharing of the positive charge of the metal ion with donor groups of the ligands. This increases the delocalization of π -electrons over the whole chelate ring and enhances the lipophilicity of the metal complexes. The increased lipophilicity enhances the penetration of the metal complexes through lipid membranes thereby blocking the metal binding sites in the enzymes of microorganisms. In addition, it disturbs the respiration process of the cell and blocks the synthesis of the proteins, thereby, restricts further growth of the organism (Dharamraj *et al.*, 2001). Methanolic leaf extract was most active against *Bacillus subtilis* and did not show any effective activity against *Staphylococcus aureus*. The results clearly showed that antibacterial activity of nanoparticles cannot be credited to the nature of plant extract and $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ individually as the LI-CuNPs showed greater activity compared to them. Moreover, the antibacterial activity of the copper nanoparticles also be attributed to presence of some phytochemical constituents of the leaf extract (Elemike *et al.*, 2017). The antibacterial activity result was shown in Table 1.

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The antibacterial potential of the methanolic leaf extract and copper nanoparticles were further confirmed at different concentrations in order to determine the complete inhibition of the samples for the bacterial strains by antibacterial kill kinetic assay. **Figure 3** showed the antibacterial kill kinetics of copper nanoparticles against *B. subtilis* and *S. aureus*.

Table 1: Antibacterial activities of Methanolic leaf extract & Copper nanoparticles

S. No.	Organism	Concentration of the sample ($\mu\text{L}/\text{well}$)	Methanolic leaf extract ($100\mu\text{g}/\text{mL}$)	CuNPs ($100\mu\text{g}/\text{mL}$)
1.	<i>Bacillus subtilis</i>	5	9.66 \pm 0.57	18.33 \pm 0.57
		10	15.66 \pm 0.57	23 \pm 1.00
		15	24.33 \pm 1.15	25 \pm 1.00
		20	31 \pm 1.00	29.66 \pm 0.47
		25	35 \pm 1.00	34.66 \pm 0.577
		50	44 \pm 1.73	46.33 \pm 1.52
2.	<i>Staphylococcus aureus</i>	5	-	7.66 \pm 0.577
		10	-	14 \pm 1.73
		15	-	20.33 \pm 0.577
		20	-	22 \pm 1.73
		25	-	29.33 \pm 0.94
		50	-	35.33 \pm 0.57

Note: Diameter (mm) of zone of inhibition (mean \pm S.D.)

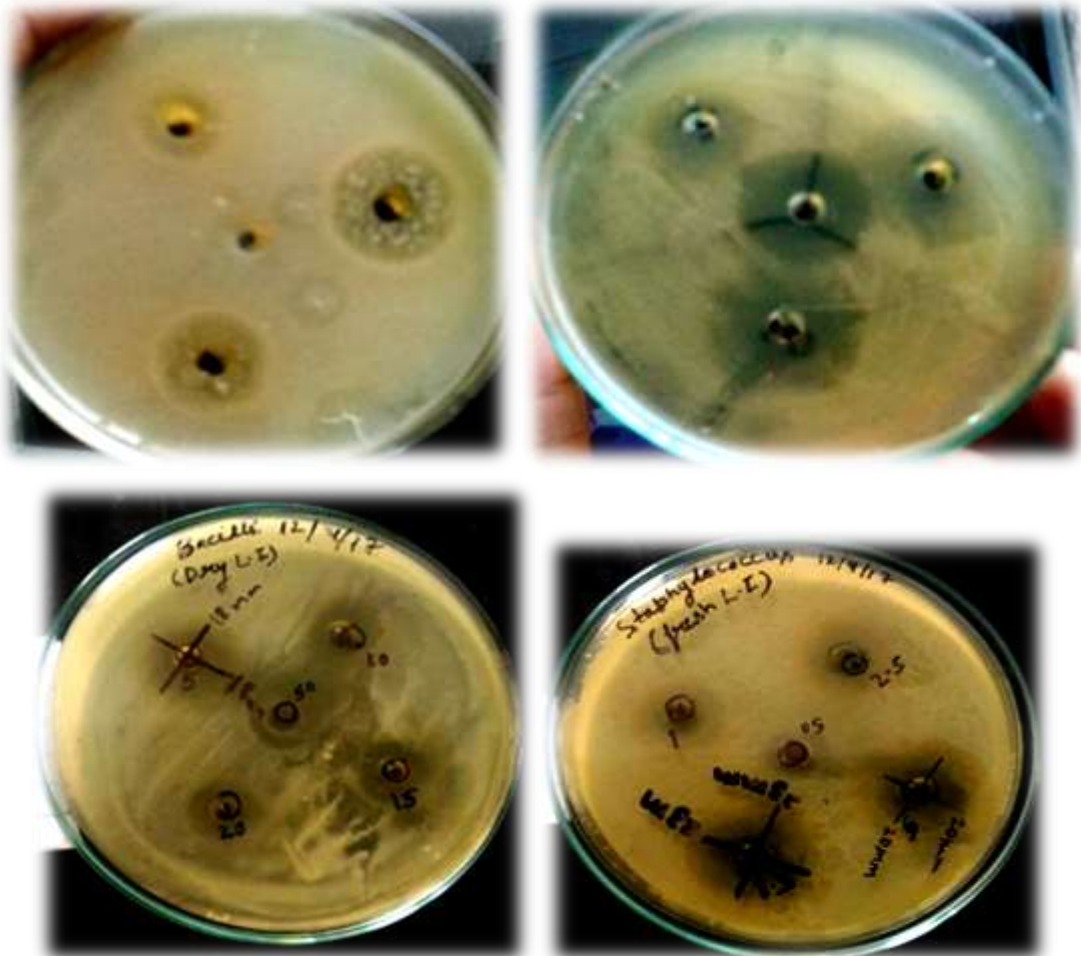


Figure 3: Antibacterial effect of Methanolic extract & CuNPs

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Plate A- showing antibacterial effect of methanolic extract of leaf against *Bacillus subtilis* at 5, 10 & 15 μL along with negative control DMSO (50 μL).

Plate B - Showing antibacterial effect of methanolic extract against *B. subtilis* at 20, 25 and 50 μL along with positive control Streptomycin (4mg/mL).

Plate C - Showing antibacterial effect of aqueous leaf extract mediated copper nanoparticles against *B. subtilis* at 5, 10, 15 and 20 μL .

Plate D- Showing antibacterial effect of CuNPs against *Staphylococcus aureus* at 5, 10, 15 $\mu\text{g/mL}$.

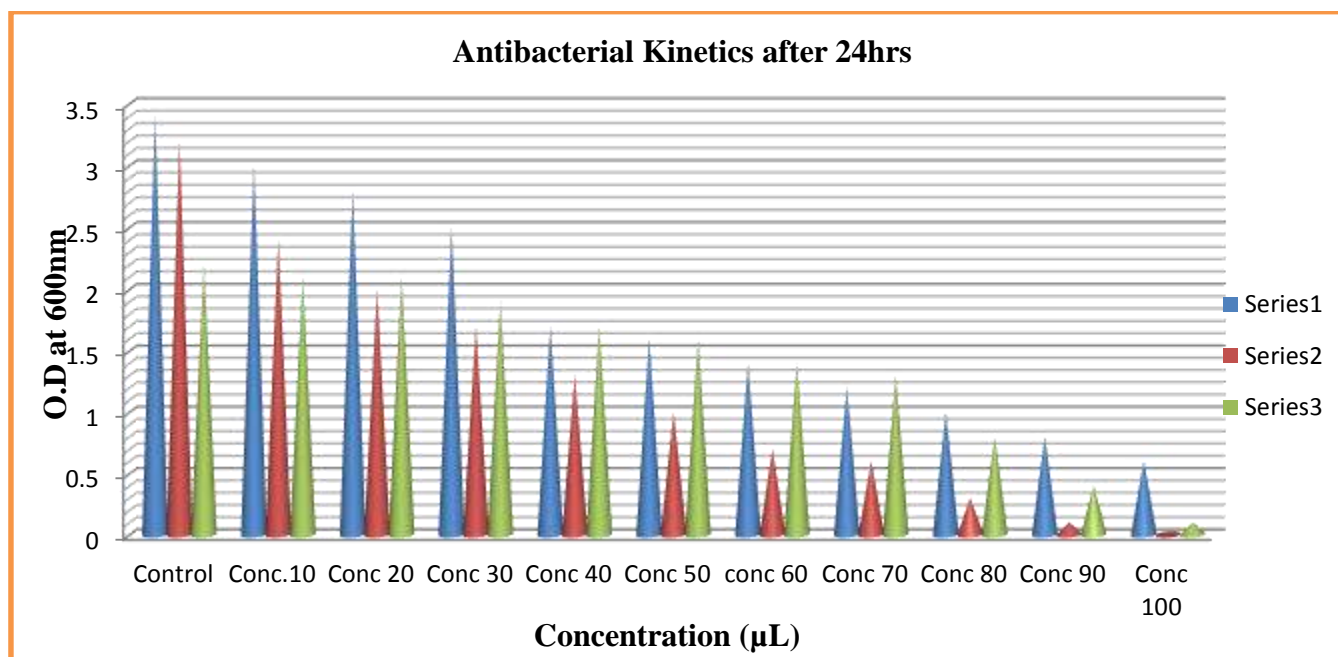


Figure 4: Antibacterial Kinetics after 24 hrs incubation

Series 1: Methanolic leaf extracts (*B. subtilis*)

Series 2: CuNPs (*B. subtilis*)

Series 3: CuNPs (*S. aureus*)

3.3 Antifungal activity (Disc Diffusion method)

Methanolic leaf extract was found efficient to inhibit fungus *C. albicans* while it did not show any activity against *A. flavus*. Whereas, the aqueous leaf extract mediated copper nanoparticles displayed antifungal activity toward the tested pathogenic strains of *A. flavus* and *C. albicans*, respectively. *A. flavus* depicted the highest sensitivity to nanoparticles compared to the *C. albicans* and was more adversely affected by the copper nanoparticles. These revealed that both methanolic leaf extract and copper nanoparticles have potential to inhibit pathogenic fungi but photosynthesized copper nanoparticles had more influence to reduce the microorganisms.

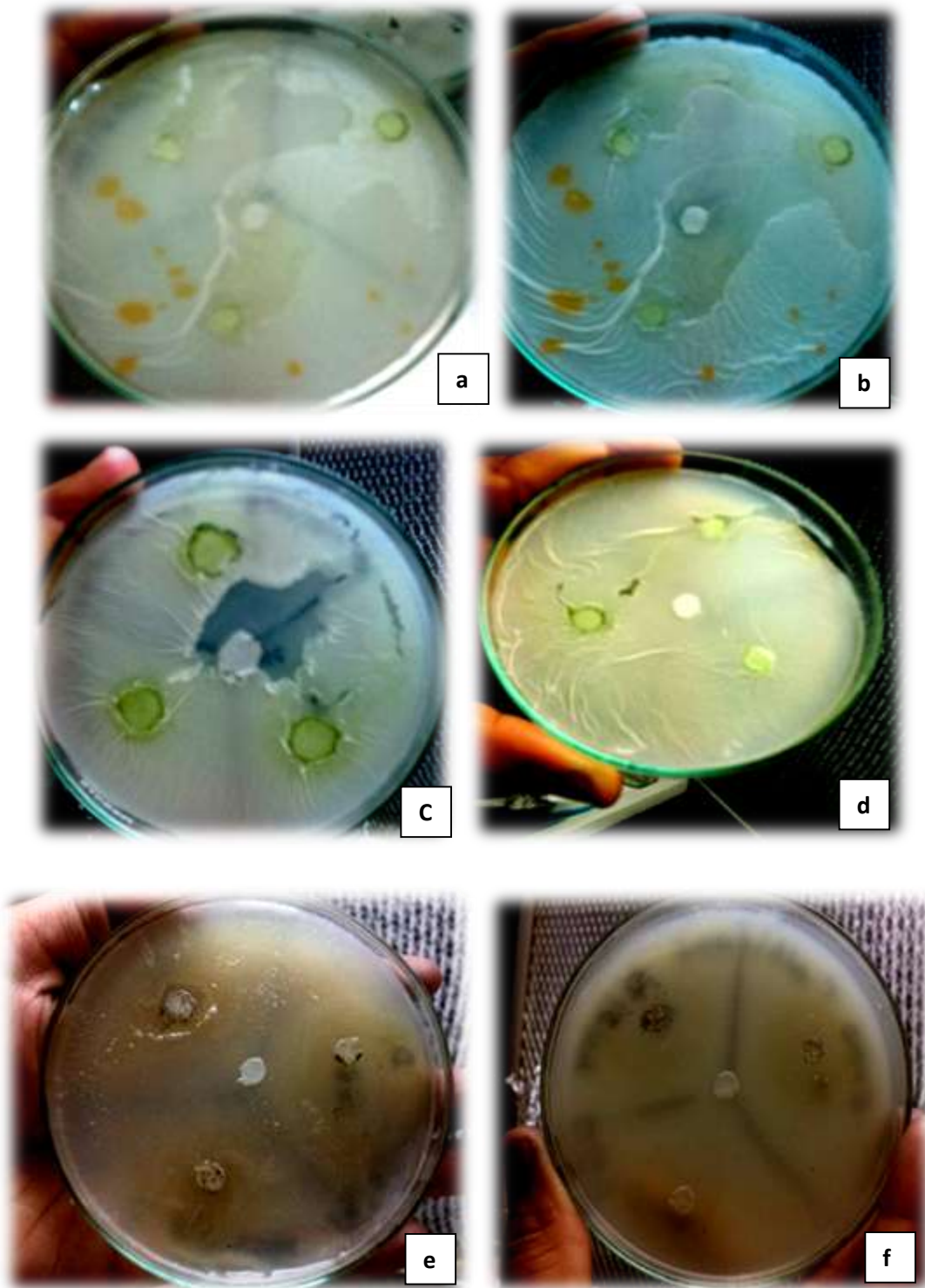
Table 4: Antifungal activity of Methanolic leaf extract and CuNPs

S.No.	Organism	Concentration of the sample ($\mu\text{L/Disc}$)	Zone of inhibition (mm) Mean \pm S.D.	
			Methanolic extract	CuNPs
1.	<i>Aspergillus flavus</i>	5	-	17.66 \pm 0.57
		10	-	26.66 \pm 0.57
		15	-	29.66 \pm 0.57
		20	-	35.33 \pm 0.57
		25	-	37.66 \pm 0.57
		50	-	42.66 \pm 0.57

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2.	<i>Candida albicans</i>	5	9±1	1±0
		10	13.66±1.52	12.66±0.57
		15	15.66±0.57	22.66±0.57
		20	19.66±0.57	29.66±0.57
		25	24.33±0.57	31±1
		50	35.33±0.57	35.33±0.57

Note: Diameter (mm) of zone of inhibition (mean ±S.D.)



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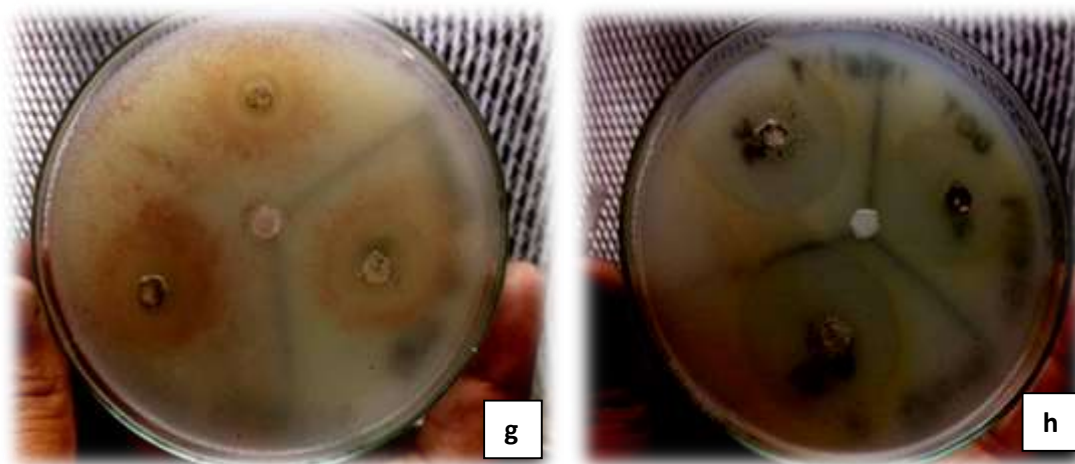


Figure 5: Antifungal effect of Methanolic extract & CuNPs

Plate a & b - Showing antifungal effects of methanolic leaf extract against *Candida albicans*

Plate c & d- Showing methanolic extract inhibition effect against *Aspergillus flavus*, there was no inhibition zone found around the extract but positive control (Streptomycin) inhibited test fungi.

Plate e & f- Showing antifungal activity of copper nanoparticles against *C. albicans* at different concentrations.

Plate g & h- Showing antifungal effect of copper nanoparticles against *A. flavus* at different concentration.

Antioxidant activity

Antioxidant activity refers to the inhibition of oxidation of molecules by inhibiting the initiation step of the oxidative chain reaction and formation of stable radicals, which are non-reactive. Plant contains phytochemicals, vitamins and other nutrients, which possesses strong antioxidant activity and helped in protecting cells against oxidative stress caused by free radicals (Mittle *et al.*, 2014). Termination of free radicals in cells prevents the pathologically disorders like cancer, heart attacks etc. (Kumar *et al.*, 2016). Antioxidant activity of both methanolic leaf extract and copper nanoparticles were carried out by DPPH assay by using ascorbic acid as standard. DPPH is a purple colored stable radical of organic nitrogen with a maximum absorbance at 517 nm and it is widely used to study radical scavenging activities of extracts and pure compounds. When odd electron becomes paired off in the presence of a free radical scavenger to form hydrazine, the absorption reduces and the DPPH solution is decolorized from deep violet to light yellow. The degree of reduction in absorbance measurement is indicative of the radical scavenging (antioxidant) power of the methanolic extract and copper nanoparticles (Pham *et al.*, 2008).

From the result, the copper nanoparticles found to be more antioxidant capacity (67%) rather than plant methanolic leaf extract (28.82%) and standard ascorbic acid (44.85%). The results signified that copper nanoparticles can be effective antioxidant agent. It is predicted that higher antioxidant activity of nanoparticles is might be due to encapsulation of bioactive molecules on the surface of CuNPs through the electrostatic attraction between negatively charged bioactive compounds (COO^- , O^-) and neutral or positively charged nanoparticles (Du *et al.*, 2013; Kumar *et al.*, 2016).

Table 5: Percentage inhibition of DPPH free radical scavenging

Sample	Concentration of sample (mg/mL)	% Inhibition of DPPH free radical scavenging activity at 517 nm
Reference	0.2	44.85
MeOH extract	0.1	28.82
CuNPs	0.1	67

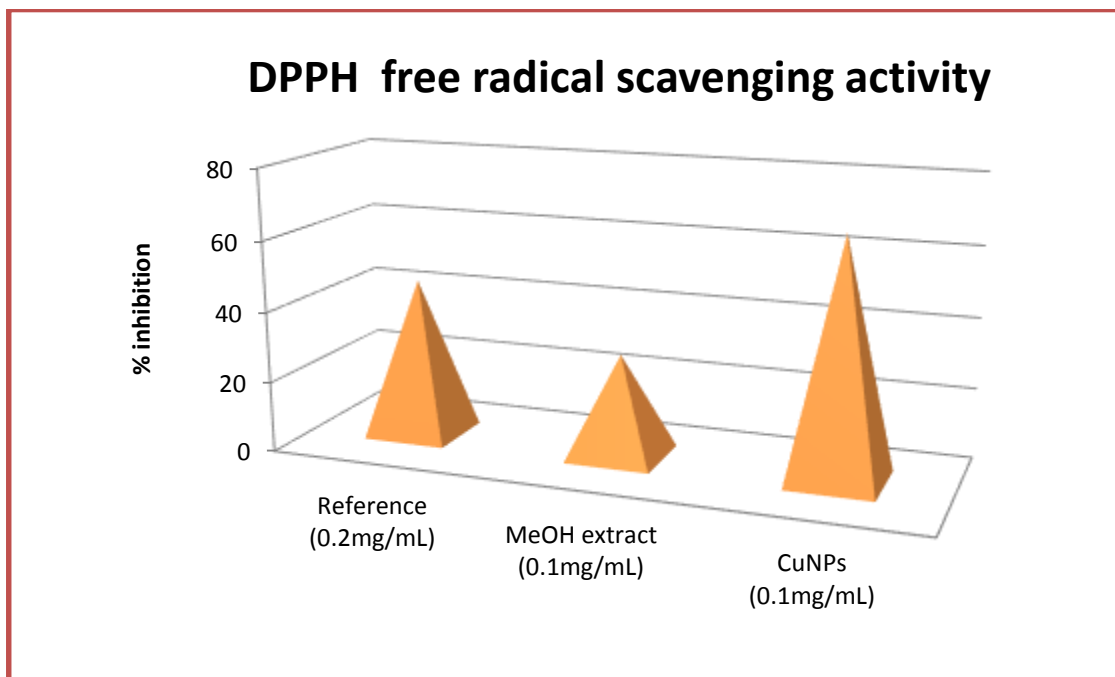


Figure 6: Percentage inhibition of antioxidant capacity

Cytotoxic activity of copper nanoparticles

Mouse macrophages cell line culture was used to demonstrate copper nanoparticles cytotoxicity as measured by the MTT assay. It is worth emphasizing that MTT assay reflects only the changes in mitochondrial function, and is not indicative of the manner or stages of cell death (Ahmad *et al.*, 2010; Rocchetta *et al.*, 2006; Cui Y *et al.*, 2014; Dan *et al.*, 2013; Qi *et al.*, 2013; Vera and Migaud, 2009; Singh 2007 and Niska *et al.*, 2015). Niska *et al.* (2015) reported that a reduction in the amount of formazon produced MTT can be proportional to the cell number. In the present study, the proportional of viable cells declined after copper nanoparticles exposure compared to control or standard.

The results of MTT assay from test sample showed different mortality rate at varying concentration. The mortality rate of mouse macrophages was found to increase with the raising of the copper nanoparticles concentration. The median cytotoxic concentration (CC₅₀) was calculated. The CC₅₀ value of copper nanoparticles was found to be 9.26054 µg/mL. So, it is evident that copper nanoparticles have greater cytotoxicity compared to other metal nanoparticles (Heinlaan *et al.*, 2008; Karlsson *et al.*, 2009). From the results, it proposes that cellular self digestion or autophagy may be involved in copper nanoparticles induced cytotoxicity. It is conserved mechanism involved in the degradation of protein and organelles in the cytoplasm (Baehrecke, 2005) or ROS generation has been proposed as possible mechanism involved in autophagy induced by copper nanoparticles (Ahmad *et al.*, 2010). Valko *et al.* (2007) and Paz-Elizur *et al.* (2008) reported that ROS considered as cytotoxic because the free radicals cause oxidative damage to biomolecules such as DNA, proteins and lipids through oxidative modifications. However, studies on the exact mechanism by which nanoparticles generated ROS in cells are still underway (Heinlaan *et al.*, 2008).

Table 3.4: Cytotoxic activity of CuNPs

S.No.	CuNPs Conc. (µg/mL)	% mortality	CC ₅₀
1	100	0.07	
2.	50	0.01	
3.	25	0.18	
4.	12.50	0.38	9.26054
5.	6.25	0.66	
6.	3.13	1.03	
7.	1.56	1.04	

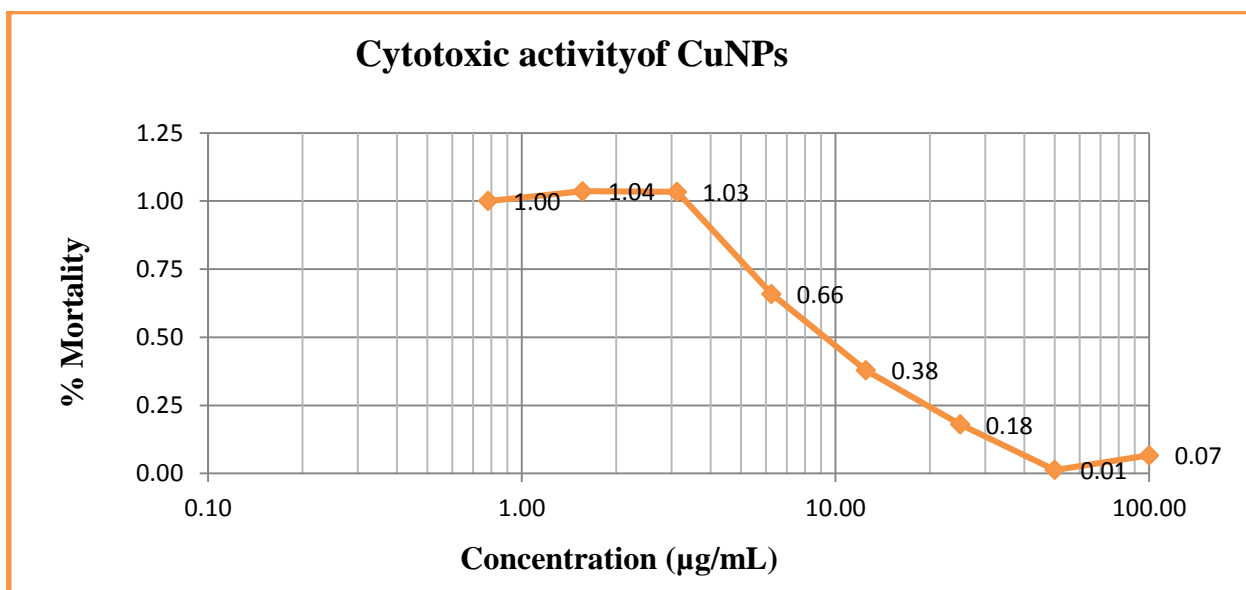


Figure 7: Showing decreasing growth rate of cell with increasing concentration of CuNPs

4. BIOCHEMICAL STUDIES

Total Protein

The leaves of Parthenium plant have wide range of proteins. The leaf extract was estimated for the water soluble proteins by using Lowry's method. The total protein was estimated from standard graph of protein and the leaf protein concentration was calculated. The calculated concentration was found to be 1.55mg/gm in leaf sample of parthenium plant. **Dickson et al. (2014)** reported the protein content in Parthenium plant was 1.75% in Parthenium spp.

Haruna et al. (2015) observed a high level of protein (24.85%) in the leaf of Parthenium plant and suggested that it could be used as a source of protein in animal diets therefore the level of protein in the leaf of Parthenium plant compare favorably well with some proteinous plants such as cowpeas (25%) (**Gallup and Reder, 1943**), pigeon pea (20.4%) (**Jambunathan et al., 1984**), lima bean, bambara groundnut (23 – 26%) (**Aletor and Aladetimi, 1989; Ene-Obong and Carnovale, 1992; Olaofe et al., 1993**).

Total Carbohydrate

The leaves of Parthenium spp. contain carbohydrate, which was estimated by using total carbohydrate method (anthrone method). The calculated value obtained from the standard graph of carbohydrate was 0.3mg/gm. **Haruna et al. (2015)** reported that crude fiber (16.41%) in Parthenium spp. which help to maintain the motility of food through the gut and may broken down by some bacteria in the gut to provide energy.

Feresin et al., (2000) described the antibacterial activity against *Staphylococcus aureus*, *S. cohnii*, *Salmonella sp.*, *Escherichia coli*, *Serratia sp.*, and *Bacillus subtilis*.

Bajwa et al., (2003) evaluated the efficacy of aqueous leaf extract of allelopathic weed, Parthenium against pathogenic fungi *Drechslera tetramera*, *Aspergillus niger* and *Phoma glomerata*.

Tapwal et al., (2011) demonstrated the antifungal activity of aqueous extract of *P. hysterophorus* against *Alternaria solani*, *A. zinniae*, *Curvularia lunata*, *Rhizoctonia solani* and *Fusarium oxysporum*.

Ahmed et al., (2016) reported that DNA isolation from the weed *P. hysterophorus* is complicated due to the presence of high amount of allelochemicals in the form of secondary metabolites that causes hindrance in extraction and enzymatic reactions.

Patel, S. (2011) demonstrated that parthenin obtained from *P. hysterophorus* is known for its allelopathic properties and for being the most toxic constituent of the plant.

Swaminathan et al., (1990), Auld and Medd, (1987) confirmed that the allelopathic potential of parthenium weed results from the release of phytotoxic substances such as, ferulic, caffeic, vanillic, chlorogenic, p-coumaric and p-hydroxybenzoic acids, parthenin, ambrosin and coronopilin, which inhibit the germination and growth of several crop plants and multi-purpose trees and also cause allergic eczematous contact dermatitis and respiratory problems in humans and livestock.

Singh et al., (2001) described some of the important weeds exhibiting allelopathy and crop losses including *P. hysterophorus*. **Kaur et al., (2014)** reviewed the effects and management of *P. hysterophorus*.

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Chung *et al.*, (2016) reported the development of a method to synthesised copper nanoparticles by using copper acetate and leaf extract of *Eclipta prostrata*. Copper nanoparticles were characterized by XRD and FTIR.

Hase *et al.*, (2016) published that Nanotechnology has wide application in various fields including Medical field. Copper nanoparticles from *Leucas chinensis* L. synthesized by green chemistry have shown antipyretics, antibacterial, antioxidant and antifungal activity. This process is environment friendly and non-toxic.

Thandapani *et al.*, (2017) established the enhanced larvicidal, antibacterial, and photo catalytic efficacy of TiO₂ nano hybrids green synthesized using the aqueous leaf extract of *P. hysterophorus*.

Anwar *et al.*, (2015) confirmed the in vitro antibacterial activity of Ag nanoparticles synthesized from leaf extract of *P. hysterophorus* L.

Rajiv *et al.*, (2013) described the bio-fabrication of zinc oxide nanoparticles using leaf extract of *P. hysterophorus* L. and its size-dependent antifungal activity against plant fungal pathogens.

Kaur *et al.*, (2017) demonstrated the phenological behaviour of *P. hysterophorus* in response to climatic variations of Chandigarh, India according to the extended BBCH scale.

Navie *et al.*, (1996) reported that Parthenium represented high genetic variability and have high invasive potential which is confirmed by statistical analysis.

Pandey *et al.*, (2003) and Chen *et al.*, (2010) evidenced that Parthenium reproduces generally by intra specific hybridization (cross pollination) and its pollens spread by amphibious mode of pollination which is the source of increased heterogeneity.

Jabeen *et al.*, (2015) conducted a study to determine the genetic structure of 95 individual samples from 11 populations (9 from Pakistan and 2 from Australia) of Parthenium using ISSR fingerprinting and results demonstrated that weed in Pakistan is genetically heterogeneous and may have been the result of multiple introductions.

5. CONCLUSION

The current study describes a method for the biological synthesis of copper nanoparticles from weed, *Parthenium hysterophorus*, which is rapid, simple, efficient, environmental friendly and non hazardous. We reported here the excellent inhibitory potential of the CuNPs against test microbial strains which are considered to be significant pharmacological targets for the treatment of such microbial infections. The potency of the biogenic copper nanoparticles for radical scavenging may be used in cosmetics to prevent aging symptoms. MTT analysis indicated that copper nanoparticles are effective against the J744 A.1 cell lines, which concluded that copper nanoparticles may use in the treatment of parasitic infections.

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